



Yield Enhancement Network

Bean YEN Welcome Pack 2026 UK

Thank you for participating in the Bean YEN 2026.

This guide provides the information you need to complete your 2026 sample submissions and how to carry out the crop observations. The samples are posted to the analytical labs throughout the season, and the crop observation and agronomy data is submitted via the YEN portal (Please note that for NCS project Pulse Pioneers, data submission will be through the BOFIN app rather than the YEN portal). We have included a progress sheet to track sampling and to make notes of the observations throughout the season. We will contact you about submitting data when the online YEN portal opens for the season. For further information or any questions please get in touch at yen@adas.co.uk.

You will receive a separate Welcome Pack for each YEN entry that you have.

Welcome Pack contents

- **Soil Sample:** Sampling instructions on page 2. Collect a representative soil sample of **~600g** from across the whole field or area to be entered in YEN and send to NRM using the labelled sample bag and **BLACK NRM pre-paid postage bag**. Include the enclosed **analysis request form** with your sample and apply the **grey NRM soil analysis label** to this.
- **Leaf Samples:** At the **start of 4th node - start of nodulation (GS 34)** send your first leaf sample to Lancrop. Send the second leaf sample at **1st flower seen sporadically within the crop (GS 60)**. Include the corresponding analysis request form with each sample and complete this with the type of crop entered, the rest of the details on these forms are prepopulated for your entry but please amend if any details are incorrect. Labelled sample bags and pre-paid postage bags are provided for these samples. Instructions on how to take tissue samples can be found at the below link:
<https://www.yara.co.uk/siteassets/crop-nutrition/farmers-toolbox/analysis/how-to-take-a-sample-for-soil-or-tissue-analysis.pdf/>
- **Grab Sample:** Instructions are on page 4. A day or two before harvest, collect **~25 whole plants (please note to collect 25 plants rather than 25 shoots)** from the YEN entry area and send in the **potato sack** provided, with the **WHITE** return address label, to ADAS Gleadthorpe via your local Post Office.
- **Return of samples:** The enclosed address labels provide **pre-paid returns** for the **grab sample**; you will not be charged when using these labels. Alternatively, you can arrange for a courier to collect the samples, which should be sent to the address mentioned at the end of this guide, however **you will not be reimbursed** for using courier services.
- **Seed Samples:** Instructions on page 5. At harvest, collect a representative sample of seed from all trailer loads from the area entered, fill the seed sample bags provided, and send one bag to each of the two laboratories for analysis, using the appropriate coloured labels and return pre-paid packaging.
 - Use the **BLUE** labels for Lancrop nutrient analysis (~200-300g)
 - Use the **GREEN** labels for PGRO Bruchid beetle and TSW analysis (~500g)
- **Yield Data:** Attach the **YELLOW** label to your Yield Entry Form (attached at the end of this guide), and record the harvest area, fresh weight, moisture content and harvest losses. Then take photos of (or scan) the completed Yield Entry form, along with your map and weighbridge tickets (if applicable) and email it to yen@adas.co.uk with your YEN ID in the subject line.

SOIL SAMPLE INSTRUCTIONS

A soil sample of the field or area to be entered in YEN should ideally be taken before any organic manure or inorganic fertiliser applications have been made. **If it is necessary to sample following an application a minimum period of 3 weeks should be allowed to pass** before doing so and details of any application(s) made provided on the analysis request form.

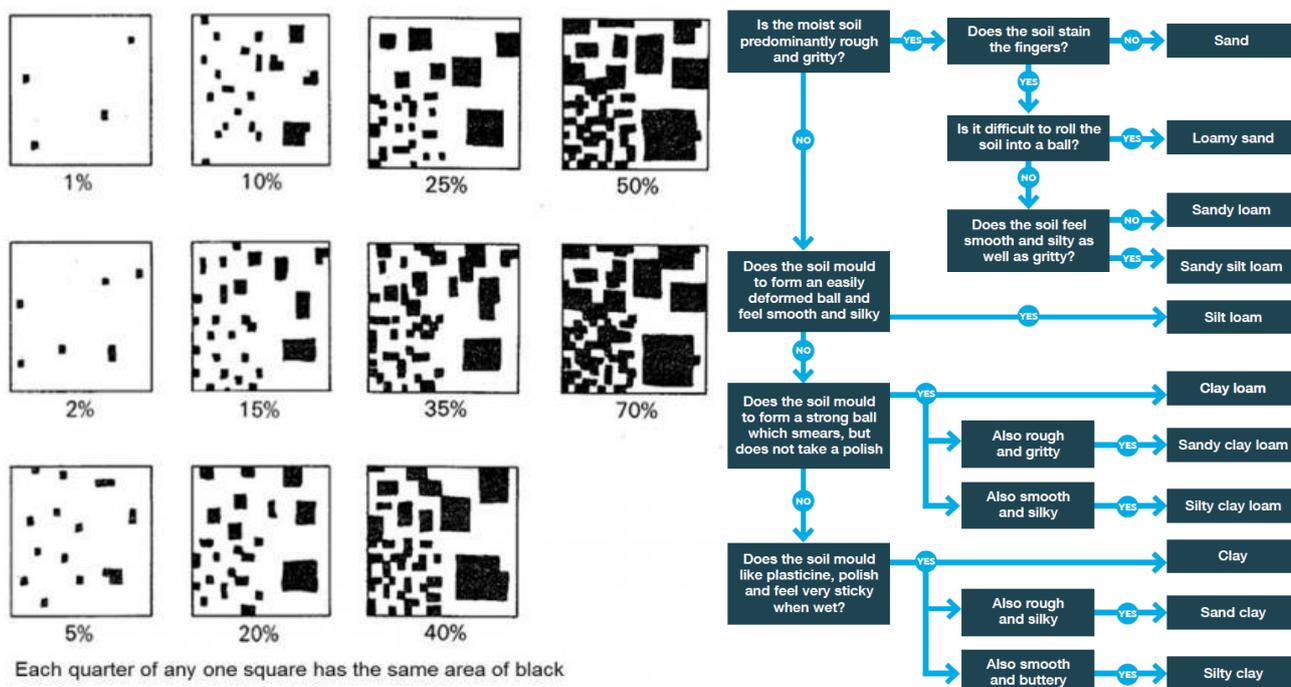
Please complete details on the form of current and previous cropping and whether previous crop residues have been removed or returned to the soil, if the field has a history of regular organic manure applications, please also detail this. **The more information you provide with your sample the better the analysis NRM will be able to complete** ensuring more reliable, trustworthy and useful results.

Walk a **'W' pattern** across the sample area as in the figure below avoiding any irregular patches such as gateways, headlands, and trees, collect about **25-30 soil cores to 15cm depth** and bulk the cores together in the labelled sample bag provided to give a **~600g sample**. Place the soil sample and analysis form with grey sticker attached into the **BLACK NRM return postage bag** and drop it off at your local post office.



When taking your soil sample please also **record the texture and approximate % stone content of the topsoil** (guidelines for assessing soil texture by hand and estimating stone content are given below). These details should be provided when completing your field and crop details form online, information about which will be emailed to you separately.

Good soil descriptions are vital in allowing us to estimate soil water holding capacity which along with rainfall data we use to determine available water for your crop, a key component in calculating yield potential.



The results of soil analysis carried out by NRM on this sample will be included in your annual report. These are also used to calculate various other metrics in the report, including available crop nutrients and efficiency of nutrient uptake.

OBSERVATION VISIT: (GS10) FULL EMERGENCE

- Record the date of full emergence (GS 10). If this is missed please record the growth stage on the date you visit.
- Carry out plant counts.
 - In at least five locations, using a meter stick or quadrat count the number of plants in a 1 m² area. If it's easier, use a 0.25 m² quadrat to count the number of plants and multiply that number by 4 to get the number of plants in 1 square metre. Please record the count in each individual location. (If your crop is a winter crop and has progressed past making an easy plant count you can count the stubble stems in a 1m² area just after harvest).
 - **Alternatively**, plant counts can be calculated from photographs. Take images at three locations from within the 2ha area entered in the YEN. Take the photo from above the crop looking vertically down, showing as wide an area as possible and including an A4 piece of paper flat on the ground for scale. See examples in Figure 1. Photos will be able to be uploaded using the online data submission portal.



○ Figure 1. Example photographs for plant population estimate

OBSERVATION VISIT: (GS 34) START OF NODULATION, FOURTH NODE (APR/MAY)

- Record date of 4th node - start of nodulation (**GS 34**)
- Collect a representative **leaf tissue sample** as below

LEAF TISSUE TESTING INSTRUCTIONS

As part of Bean YEN Lancrop/YARA provide free tissue testing for YEN entrants at the start of 4th node - start of nodulation (GS 34) and at 1st flower seen sporadically within the crop (GS 60).



Include the corresponding analysis request form with each sample and complete this with the type of crop entered, the rest of the details on these forms are prepopulated for your entry but please amend if any details are incorrect.

At each sampling timing:

- Sample and send Monday to Wednesday to avoid the sample being in the post over the weekend.
- Within your YEN area walk up 2 to 4 representative tramlines and sample tissue at regular intervals from between 5 - 20 points along the sampling path.
- At each sampling site select several plants at the same stage of development and sample the youngest mature leaf without the petiole (first fully expanded leaves away from the growing point) until you have between 300g – 400g of material.
- Avoid leaves showing pest, disease or other damage. Take leaves only, not stems.

- Mix the leaves thoroughly, if wet blot the leaves dry with a paper towel and place into a sample bag, squeezing out the excess air and sealing.
- If foliar nutrition is to be applied to the crop at flowering, please take the second tissue sample before any flowering foliar nutrient sprays are applied.
- Fill in the order form including crop and growth stage. Include your email to ensure you get the results.
- Place the sample bag and the order form into a Lancrop/Yara pre-paid envelope and post. Do not put the order form inside the bag with the sample as it may get wet.

NODULATION SCORES: (GS50) FIRST FLOWER BUDS (APRIL/MAY)

- This is an optional assessment for those interested in understanding their crop's nodulation.
- The main assessment timing is when the first flower buds are forming (BBCH GS50). Later or earlier assessments could provide information on when nodulation is likely to begin and end.
- Walk a 'W' pattern across the field or treatment area avoiding any irregular patches such as gateways, headlands, and trees, and collect material from at least four different areas.
- At each area dig out aim to dig up one plant, with a relatively undisturbed root crown. Dig up that plant using a spade. Start digging 10-15 cm away from the selected plant to prevent damaging the root crown. Insert the spade to its full depth ~20 cm around the plant and dig up the soil and the plant. You may end up digging up a few plants depending on your seed rate. Allow any soil that easily falls away to do so. Carefully remove any excess soil that easily falls away.
- Soak the plant root systems and accompanying soil in a bucket of water and a small amount of soap for at least 15 minutes to loosen the soil from the roots. Gently move the root systems back and forth to assist soil removal. Carefully wash the majority of the soil from the roots, being careful not to knock the nodules from the roots.
- Once the root system is relatively free from soil, use the score chart on the next page to score nodulation and check if the nodules are active by cutting the nodules open with a sharp knife and seeing if they turn a pinkish/red colour
- Complete the scoring sheet at the end of this guidance, take a picture of the scoring sheet and send it to yen@adas.co.uk

OBSERVATION VISIT: (GS60) FIRST FLOWER (MAY/JUNE)

- Record date of 1st flower seen sporadically within the crop (**GS 60**).
- Collect a representative **leaf tissue sample**, following the method described above.

OBSERVATION VISIT: (GS69) END OF FLOWERING (JUNE)

- Record date when crop is out of flower (**GS 69**).

OBSERVATIONS VISIT: (GS97) FULL SENESCENCE

- Record date when crop is first ripe for harvest, full senescence (GS97)
- Take your grab sample as detailed below

GRAB SAMPLE INSTRUCTIONS

The potato sacks and plastic bag enclosed in the Harvest Pack are for use in sending whole-crop 'grab' samples and grain samples to ADAS Gleadthorpe for analysis. These samples are needed as part of your Bean YEN competition entry; they will enable us to produce a yield analysis report which will explain your crop's performance and allow us to suggest how yields might be improved in the future. It will help ensure trustworthy results if you follow these instructions carefully. Please address any questions to the named contacts below.

A grab sample of the whole crop, **including the roots**, must be taken once the crop has reached harvest maturity, when the crop is in a dry state, ideally a day or two before harvest. If the roots cannot be removed from the soil cut the plant where the stem meets the soil.

Go to the field with the two sacks and a serrated blade or secateurs.

- The one representative sample should comprise of 5 sub-samples from 5 points within your yield area totalling exactly 25 plants; we suggest the following sub-sampling procedure, as in the example to the right:
- select two typical tramlines running through the intended area for yield measurement
- estimate the number of paces which will take you the length of the area
- walk a third of this length along the first tramline, then step away from the tramline by 3-4 paces
- without close inspection, randomly select ~5 neighbouring plants and pull them at ground level to extract as much of the roots as possible.
- place the whole plant, top-first into the sack
- repeat for the next 2 points along the first tramline, and then repeat on alternate sides as you walk back along the second tramline, this time selection plants from two locations to collect 25 plants in total.

Grab samples for Research Plot trials

- For research trial yields a grab sample should consist of 7-8 shoots per plot, from a minimum of 3 replicate plots. Put shoots from all replicate plots into the same sack for despatch.

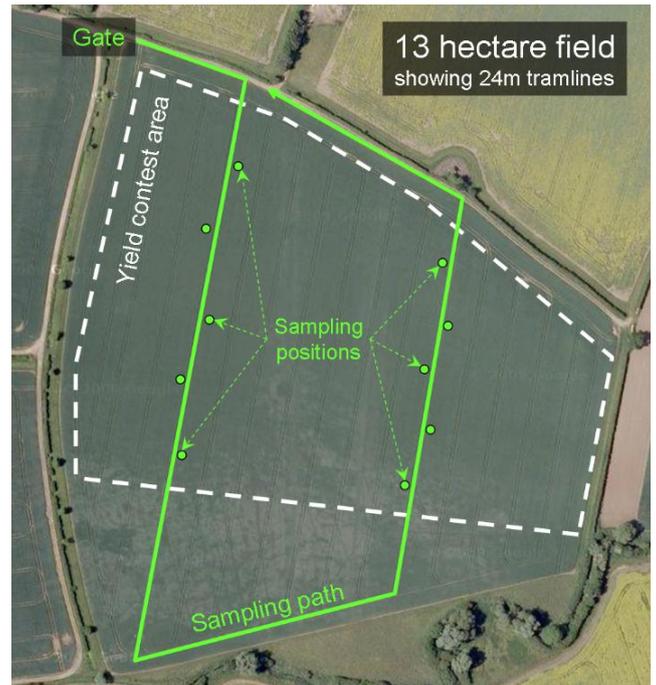
Packing your grab sample

- Place all 25 plants side-by-side, pods-first, into the sacks, and seal the end. Please note that both sacks do not need to be used if you can fit all 25 plants into one.
If any shoot ends are protruding, fold them over inside the sack so that you can seal the end, with the length being <60 cm. **NB: The final package must be no more than 60cm long.** If the package is longer than this, it may be rejected or 'lost in transit'.
- Attach a **WHITE** pre-paid postal address sticker onto the sack and take it to your local post office/pick up point to be sent to ADAS Gleadthorpe.

What will your grab sample tell you?

Data collected from your grab sample will be fed back in your end of year report. The metrics we derive from your grab sample include:

- Total Crop Biomass
- Harvest Index – the proportion of total biomass that is grain
- Pods per shoot
- Seeds per pod
- Plant height
- Estimated use of available water
- Estimated % solar radiation captured.



AT HARVEST

- At harvest record your yield as per the yield form. This can be via weigh bridge or via a calibrated combine yield mapper.
- Record harvest losses as below
- Take your seed samples as below

HARVEST LOSSES INSTRUCTIONS

Count the number of beans in an A4 paper sized area at 5 locations directly behind the combine, and at 5 locations between swaths (i.e. where beans would only be present by shattering before entering combine, not due to losses over the sieves).

Record the number of beans per area at each location using the provided Yield Entry Form.

SEED SAMPLE INSTRUCTIONS

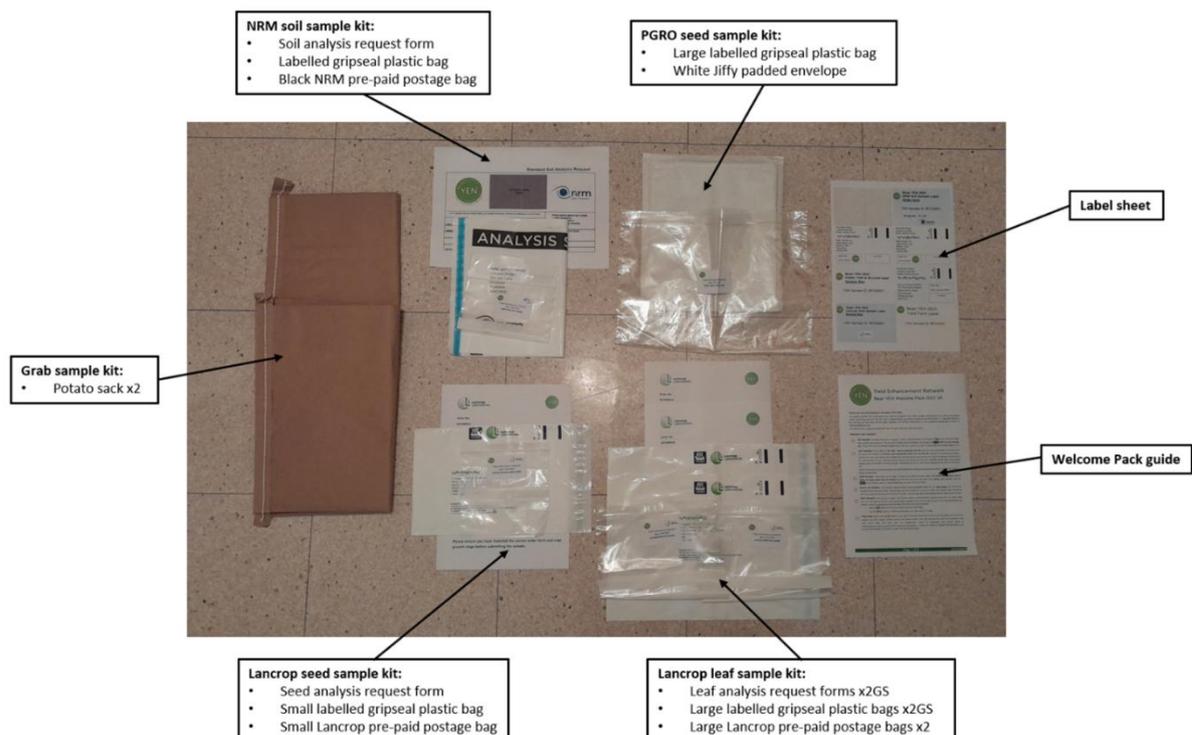
We request that all Bean YEN Entrants return TWO seed samples at harvest, each sent directly to separate laboratories for analysis.

Collecting your Grain sample

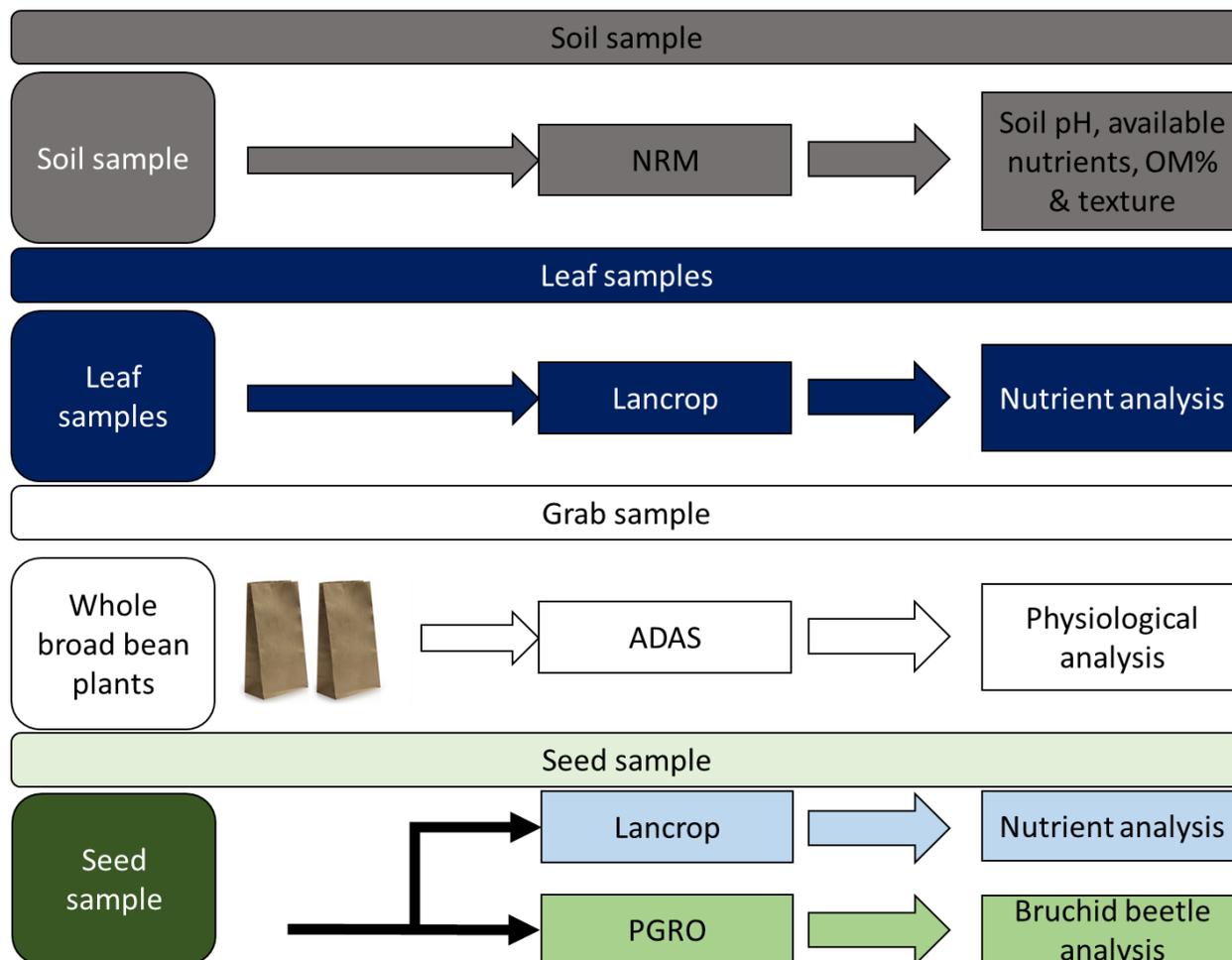
- A sample of ~800g (just under 2 lb) seed should be taken at or just after harvest. Take an appropriate size sub-sample (~200g) from each trailer-load of seed close to the time it is being weighed – for example, straight afterwards, as it is being tipped. The sample should represent the seed being weighed, so it should be taken from several parts of the trailer load and it should not be cleaned, dried or otherwise changed before it is placed in the plastic bag provided.
 - Amalgamate and thoroughly mix the sub-samples in a bucket. The composite seed sample should be divided into two sub samples:
1. Lancrop seed Sample ~200-300g
 - Place a 200-300g subsample into the Lancrop seed tissue testing zip lock bag (this should be about half full). A Lancrop order form has been generated with your details on. Check and amend if necessary and then place the Lancrop order form also inside the zip lock bag and seal. Stick the **BLUE** Lancrop seed sample label on the outside of the sealed bag, and seal the whole thing in the Lancrop self-return pre-paid envelope and return to Lancrop via your local post office.
Nutrient analysis carried out by Lancrop on this sample will be included in your annual report.
 2. PGRO Grain Sample (via ADAS) ~500g
 - Place a 500g subsample of seed into the clear plastic bag provided for PGRO, attach the **GREEN** Seed Sample bag label to the clear plastic bag, and then place the sealed seed sample into the grey mail bag also provided, and seal this. If the clear plastic bag is half full, you have provided sufficient seed. Attach the **GREEN** Seed Sample return address label to the grey mail bag and return to PGRO for analysis via your local post office.

PGRO will measure Bruchid beetle damage and TSW which will be included in your report. These are also used to calculate various other metrics in the report, including seeds/m² at harvest

WELCOME PACK CONTENT SUMMARY



WELCOME PACK POSTAGE SUMMARY



RETURN ADDRESSES

Lancrop	ADAS Gleadthorpe	NRM	PGRO
Yara Analytical Services Pocklington Industrial Estate Pocklington York YO42 1DN	FAO Bean YEN ADAS Gleadthorpe Meden Vale Mansfield NG20 9PD	Coopers Bridge Braziers Lane Winkfield Row Bracknell RG42 6NS	PGRO The Research Station Great North Road Thornhaugh Peterborough PE8 6HJ

If you require additional labels for any samples, please contact yen@adas.co.uk.

Crop samples, Yield data and Online Field Data submission forms must be returned no later than **30 September**

THE PGRO PEA AND BEAN GUIDE APP

PGRO have developed a free agronomy app, which will be useful in the field. It can aid with pest and disease recognition and is capable of recording and submitting reports of pests and diseases. There is also a built in growth stage guide. The app is available for both Android and Apple devices.

GROWTH STAGES KEY

Faba bean Weber and Bleiholder, 1990; Lancashire et al., 1991

Code Description

Principal growth stage 0: Germination

- 00 Dry seed
- 01 Beginning of seed imbibition
- 03 Seed imbibition complete
- 05 Radicle emerged from seed
- 07 Shoot emerged from seed (plumule apparent)
- 08 Shoot growing towards soil surface
- 09 Emergence: shoot emerges through soil surface

Principal growth stage 1: Leaf development¹

- 10 Pair of scale leaves visible (may be eaten or lost)
- 11 First leaf unfolded
- 12 2 leaves unfolded
- 13 3 leaves unfolded
- 1 . . . Stages continuous till . . .
- 19 9 or more leaves unfolded

Principal growth stage 2: Formation of side shoots

- 20 No side shoots
- 21 Beginning of side shoot development: first side shoot detectable
- 22 2 side shoots detectable
- 23 3 side shoots detectable
- 2 . . . Stages continuous till . . .
- 29 End of side shoot development: 9 or more side shoots detectable

Principal growth stage 3: Stem elongation

- 30 Beginning of stem elongation
- 31 One visibly extended internode²
- 32 2 visibly extended internodes
- 33 3 visibly extended internodes
- 3 . . . Stages continuous till . . .
- 39 9 or more visibly extended internodes

¹ Stem elongation may occur earlier than stage 19; in this case continue with the principal stage 3

² First internode extends from the scale leaf node to the first true leaf node

Principal growth stage 5: Inflorescence emergence

- 50 Flower buds present, still enclosed by leaves
- 51 First flower buds visible outside leaves
- 55 First individual flower buds visible outside leaves but still closed
- 59 First petals visible, many individual flower buds, still closed

Principal growth stage 6: Flowering

- 60 First flowers open
- 61 Flowers open on first raceme
- 63 Flowers open 3 racemes per plant
- 65 Full flowering: flowers open on 5 racemes per plant
- 67 Flowering declining
- 69 End of flowering

Principal growth stage 7: Development of fruit

- 70 First pods have reached final length ("flat pod")
- 71 10% of pods have reached final length
- 72 20% of pods have reached final length
- 73 30% of pods have reached final length
- 74 40% of pods have reached final length
- 75 50% of pods have reached final length
- 76 60% of pods have reached final length
- 77 70% of pods have reached final length
- 78 80% of pods have reached final length
- 79 Nearly all pods have reached final length

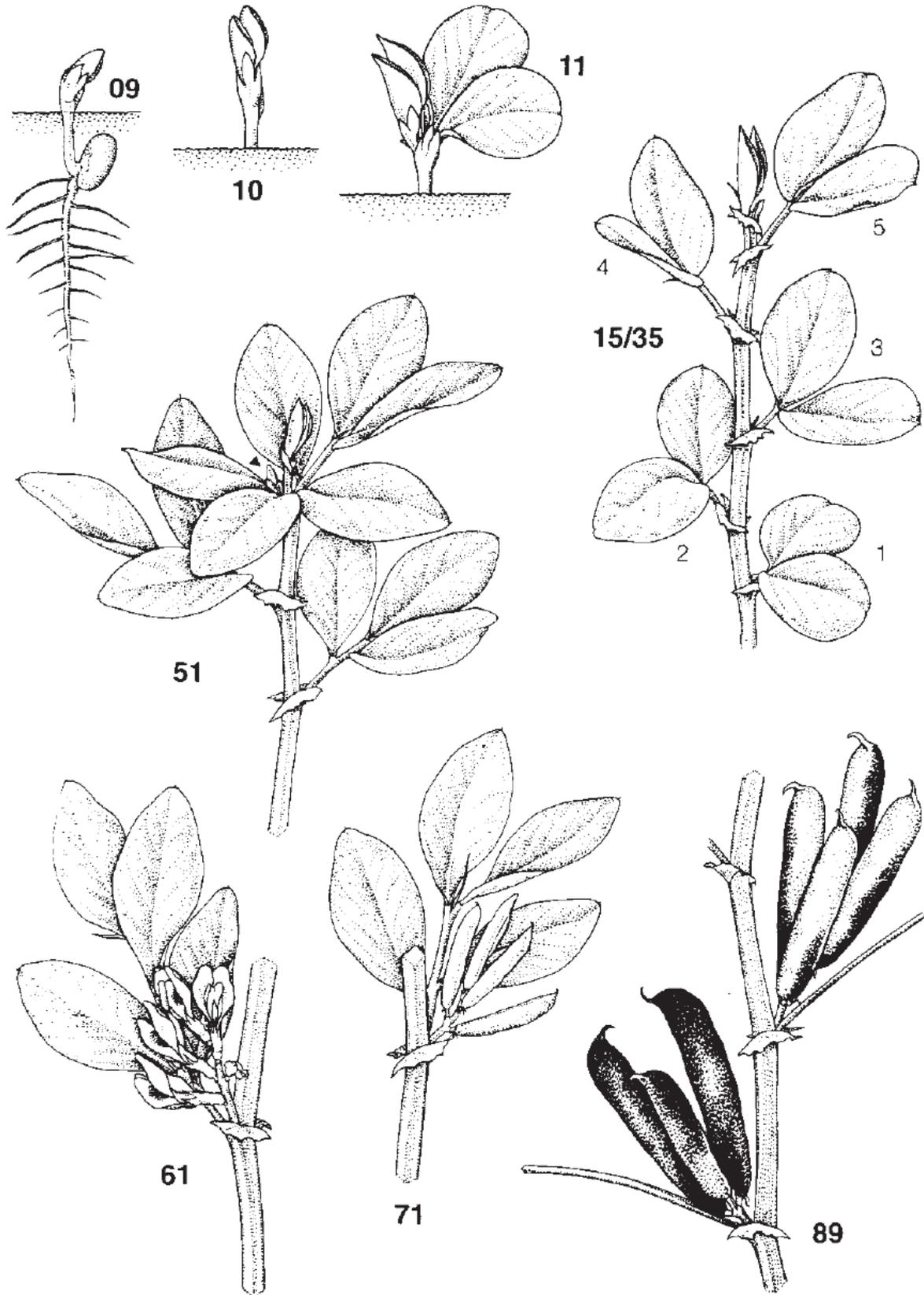
Principal growth stage 8: Ripening

- 80 Beginning of ripening: seed green, filling pod cavity
- 81 10% of pods ripe, seeds dry and hard
- 82 20% of pods ripe, seeds dry and hard
- 83 30% of pods ripe and dark, seeds dry and hard
- 84 40% of pods ripe and dark, seeds dry and hard
- 85 50% of pods ripe and dark, seeds dry and hard
- 86 60% of pods ripe and dark, seeds dry and hard
- 87 70% of pods ripe and dark, seeds dry and hard
- 88 80% of pods ripe and dark, seeds dry and hard
- 89 Fully ripe: nearly all pods dark, seeds dry and hard

Principal growth stage 9: Senescence

- 93 Stems begin to darken
- 95 50% of stems brown or black
- 97 Plant dead and dry
- 99 Harvested product

Faba Bean



CONTACTS

Thomas Wilkinson Thomas.Wilkinson@adas.co.uk 07503570264

Charlotte White Charlotte.White@adas.co.uk

Or email yen@adas.co.uk for general enquiries.

@adasYEN



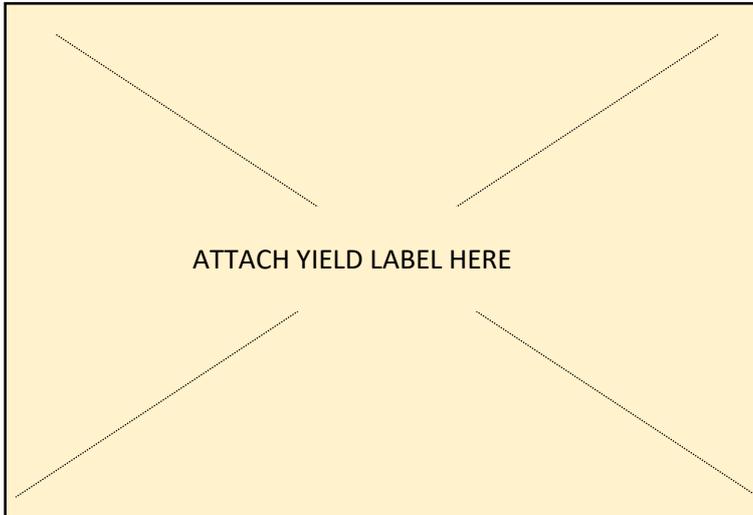
YEN SPONSORS IN 2026





Bean Yield Enhancement Network 2026

Please fill the below form and send a copy with weighbridge ticket(s) (if applicable) to yen@adas.co.uk with your YEN ID in the subject line.



Yield Entry Form

Field Name.....

Harvest area (hectares)

Fresh seed weight at harvest

Average moisture content of seed (%)

Briefly describe how you calculated your harvest area:

Location	Beans/A4 area between swaths	Beans/A4 area behind combine
1		
2		
3		
4		
5		

Have you included a weighbridge ticket? YES NO

Combine Yield Monitor/Map (if available)

Yield from combine yield monitor/map

Yield monitor make and type

GPS accuracy

Date yield monitor was last calibrated

Harvest area: I confirm that the description of the harvest area accurately represents the area entered into Bean YEN.

Harvest weight: I confirm that the harvest weight reported relates to seed harvested from the area described.

Seed sample: I confirm that the seed sample in the bags entered into the Bean YEN contain seed

Name.....

Date.....

NODULATION SCORING SHEET

0	0.5	1	2	3	4	5	6	7	8
Absent	Ineffective	Rare (Effective)	Scarce	Moderate	Adequate	Ample	Abundant	Very Abundant	Extremely Abundant
									
No nodules	White ineffective nodules	0-5 small pink nodules	6-10 small pink nodules	11-20 small pink and/or 1-2 large pink nodules	21-40 small pink and/or 3-4 large pink nodules	< 40 small pink and/or 5-9 large pink nodules	≤ 10 large pink nodules	Crown nodulation <5 mm diameter	Crown nodulation ≥ 5 mm diameter

RECORD SHEET

Pulse YEN ID (e.g Bean26000XX)			
Date of measurement (DD/MM/YY)			
Crop growth stage (BBCH)			
Further notes			
Was there signs of pea and bean weevil damage in the early spring (circle one)? See PGRO technical update 08	Yes	No	Not assessed

Position	Nodule score (1-9)	Active nodules, Yes or No?
1		
2		
3		
4		